



A Rare Case of Acute Myeloid Leukemia With *SET-NUP214* Fusion and Massive Hyperdiploidy

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Dear Editor,

The *SET-NUP214* fusion gene has been observed most commonly in T-cell ALL (T-ALL) patients, with a frequency of about 3.3% to 10.3%, and in acute undifferentiated leukemia (AUL) patients, but is rare in AML patients [1-4]. Most patients diagnosed as having T-ALL have normal karyotypes according to conventional cytogenetic analysis [2]. *SET-NUP214* fusion, though reported rarely, has a significant value in acute leukemia because it often predicts poor patient outcomes [1]. We report a rare case of AML with *SET-NUP214* fusion and massive hyperdiploidy. To the best of our knowledge, this is the first case of AML with *SET-NUP214* fusion in Korea. This study was approved by the Institutional Review Board of Dong-A University Hospital, Busan, Korea, and informed consent was obtained from the patient.

A 46-year-old Korean male visited the emergency department of Dong-A University Hospital for further evaluation of fever and general weakness. Clinical evaluations were unremarkable, except for mild haziness on a chest X-ray, suggesting pneumonia. Initial laboratory results showed mild leukocytosis ($17.1 \times 10^9/L$ leukocytes), severe anemia (53 g/L Hb), and thrombocytopenia ($120 \times 10^9/L$ platelets). Differential count revealed 89% blasts with 1% segmented neutrophils, 7% lymphocytes, and 3% monocytes. A bone marrow aspirate smear showed 88% of pleomorphic blasts characterized by large size, high nucleic/cytoplasmic

ratio, irregular nuclear membranes and convolutions, and cytoplasmic projections (Fig. 1A). A bone marrow biopsy revealed portions of marked hypercellular areas and moderate fibrosis. The blasts were negative for periodic acid-Schiff and non-specific esterase stains and mostly positive for Sudan black B and myeloperoxidase (MPO) stains. In flow-cytometric analysis, the blasts were positive for MPO, CD33, CD7, CD34, and CD71 antigens, indicating AML.

Multiplex-nested PCR with HemaVision (DNA Diagnostic A/S, Risskov, Denmark) revealed *SET-NUP214* fusion transcripts of 393 base pairs. Chromosome analysis revealed 59-90,XXXY,-1,-2,-5,-7,-7,-10,-13,-13,-16,-17,-18,-21[cp23], interpreted as a composite karyotype with massive hyperdiploidy in all of the 23 examined chromosomes, without any apparent structural abnormalities (Fig. 1B). FISH showed tetrasomy signals for the *MLL*, *ERG1*, *CEP7*, *TP53*, and *MECOM* genes. Because *SET-NUP214* rearrangement can result from either del(9)(q34.11q34.13) or t(9;9)(q34;q34) [5], further studies with additional FISH probes were conducted. Based on the results of FISH analysis with a *BCR/ABL1* probe, we suspected that the *SET-NUP214* fusion gene in this patient was probably formed by deletion of chromosome 9, because *ABL1* (9q34.12) is located between *SET* (9q34.11) and *NUP214* (9q34.13), and the two red signals were missing (Fig. 1C and 1D) [6, 7]. However, Sanger sequencing indicated

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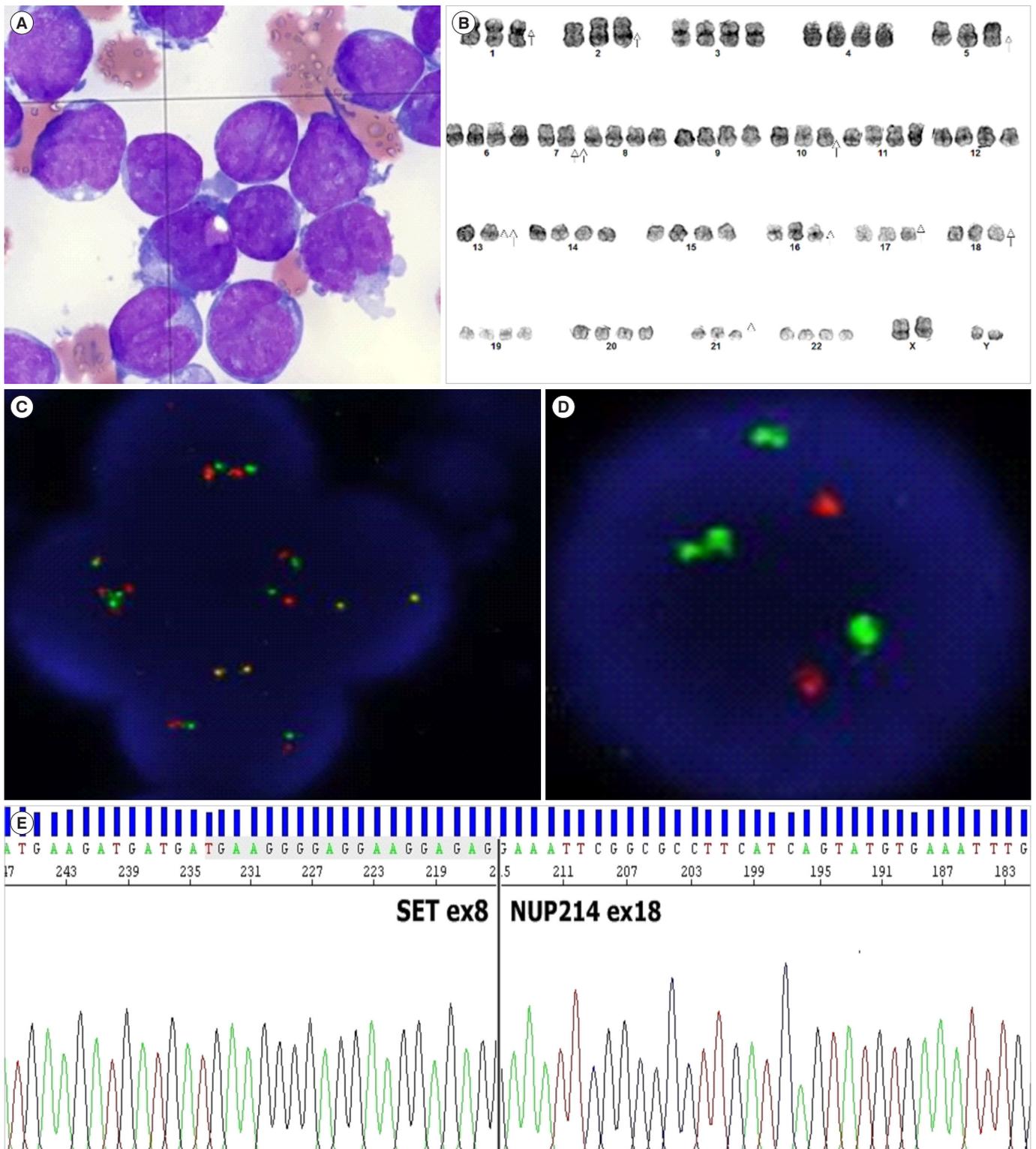


Fig. 1. Features of bone marrow aspirate smear and results of chromosomal analysis, FISH, and Sanger sequencing. (A) Bone marrow aspirate smear showing leukemic blasts comprising mostly myeloblasts (Wright-Giemsa stain, $\times 1,000$). (B) Chromosomal analysis showing 59-90,XXYY,-1,-2,-5,-7,-7,-10,-13,-13,-16,-17,-18,-21[cp23]. (C) FISH analysis with a *SET/NUP214* probe showing two yellow fusions, and two green and two red probes, suggesting either $\text{del}(9)(\text{q}34.11\text{q}34.13)$ or $\text{t}(9;9)(\text{q}34;\text{q}34)$. (D) FISH analysis with *ABL1* probe showing two red (*ABL1*) and four green (*BCR*) signals. (E) Sanger sequence analysis of *SET* and *NUP214*.

Table 1. Characteristics of a previous and the present case of AML with *SET-NUP214* fusion

No. case (N)	Sex	Age (year)	Dx.	Additional chromosomal abnormalities		Reference
1	M	35	AML-M4	None	del(9)(q34.11q34.13)	Rosati <i>et al.</i> [8]
2	M	46	AML-M1	59–90,XXXY,-1,-2,-5,-7,-7,-10,-13,-13,-16,-17,-18,-21[cp23]	t(9;9)(q34;q34)	Present case

Abbreviations: M, Male; AML-M1, acute myeloid leukemia with maturation; AML-M4, acute myelomonocytic leukemia; del, deletion; t, translocation; Dx., Diagnosis.

that the *SET-NUP214* fusion was generated by chromosomal translocation (Fig. 1E).

The patient was diagnosed as having AML without maturation with *SET-NUP214* fusion. Induction and consolidation chemotherapy with idarubicin and cytosine arabinoside was given, and in approximately three months, the patient achieved complete hematologic (2.6% of myeloblasts) and molecular remissions (normal male karyotype and negative for *SET-NUP214* fusion as indicated by PCR). After approximately five months, the patient received allogeneic peripheral blood stem cell transplantation from a full-matched sibling donor. Currently, with successful platelet and neutrophil engraftment and sufficient CD34 counts, the patient is being treated for cytomegalovirus infection.

Since *SET-NUP214* fusion was first detected in an AML patient with a normal karyotype in 2007 [2], 43 cases of acute leukemia with such rearrangement, including the current case, have been reported worldwide [1]. AML comprises only a very small portion (~2.4%) of these cases, and a hyperdiploid karyotype is also extremely rare. Characteristics of a previous [8] and the present case of AML with *SET-NUP214* fusion are presented in Table 1.

NUP214 is essential for cell cycle progression and nucleocytoplasmic transport, and *SET* is involved in inhibiting apoptosis caused by cytotoxic T lymphocytes [9]. The fusion of these genes is recurrent in T-ALL, and it is strongly associated with corticosteroid and chemotherapy resistance [10]. However, the prognostic significance of this fusion in AML with numerical chromosomal abnormalities remains unclear. We believe that such genetic rearrangements with additional chromosomes are clinically heterogeneous, and it is difficult to place the current case into a specific prognostic group. Because this genetic rearrangement is cryptic in conventional karyotyping, additional analytical methods, such as reverse transcription PCR or FISH, should be used.

Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article are reported.

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