



## First Case of *Aureimonas altamirensis* Bacteremia in Korea

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Dear Editor,

*Aureimonas altamirensis*, a gram-negative, nonmotile bacterium that grows in strictly aerobic environments, was isolated from the subterranean environment of Altamira Cave in Spain in 2006 and was designated *Aurantimonas altamirensis* [1]; however, it was reclassified as *Aureimonas altamirensis* with other *Aurantimonas* species. In 2008, it was first isolated from clinical samples [2]. *A. altamirensis* has usually been implicated as a contaminant originating from the environment, such as water sources [2], and is reported as an agent of canine infection [3]. To date, only a few cases of human infections have been reported (Table 1) [4-8]. We report the first case of bacteremia by *A. altamirensis* in Korea. The Institutional Review Board of Chonbuk National University Hospital, Jeonju, Korea, determined that this study has a low ethical load and did not require informed consent from the patient (IRB No. CUH 2018-03-028).

A 65-year-old male presented to the emergency room of Chonbuk National University Hospital because of fever and general weakness in January 2015. Five and 10 years previously, he had been diagnosed as having diabetes mellitus and a stroke, respectively, but had been well prior to admission. On initial physical examination, the patient exhibited no unusual features, and his vital signs were normal except for a body temperature of

38.9°C. His white blood cell count was  $19 \times 10^9/L$  (neutrophils 96%), Hb level was 12.3 g/L, and platelet count was  $380 \times 10^9/L$ . Total bilirubin and direct bilirubin levels were elevated at 49.26 and 43.27  $\mu\text{mol/L}$ , respectively. His AST level was 0.99  $\mu\text{kat/L}$ , ALT level was 1.00  $\mu\text{kat/L}$ , glycated Hb (HbA1c) level was 6.5%, and C-reactive protein level was 1,971.47 nmol/L in initial blood tests. Abdominal computed tomography showed unremarkable findings. Two sets of blood culture (FA Plus, FN Plus, BacT/Alert 3D system, bioMérieux, Durham, NC, USA) were done aseptically, and one of them was positive. From the subculture on blood agar plate incubated on 35°C with non-CO<sub>2</sub> condition for 24 hours, grayish colonies were observed. They were gram-negative bacilli that were catalase- and oxidase-positive. The isolate did not grow well in a 5% CO<sub>2</sub> condition; it grew well on both blood agar and MacConkey agar plates under non-CO<sub>2</sub> conditions but did not produce any colonies on MacConkey agar plates when incubated with 5% or 10% CO<sub>2</sub> for 24 hours. The Vitek 2 system (bioMérieux Inc., Hazelwood, MO, USA) identified this organism as *Acinetobacter lwoffii* (96% probability), while the Vitek MS system (bioMérieux, Marcy L'Étoile, France) could not identify the organism. 16S ribosomal RNA sequencing (GenBank, MK749933) showed 99.6% (1,361/1,366) homology with *Aureimonas altamirensis* (GenBank, NR\_043764.1), and the %

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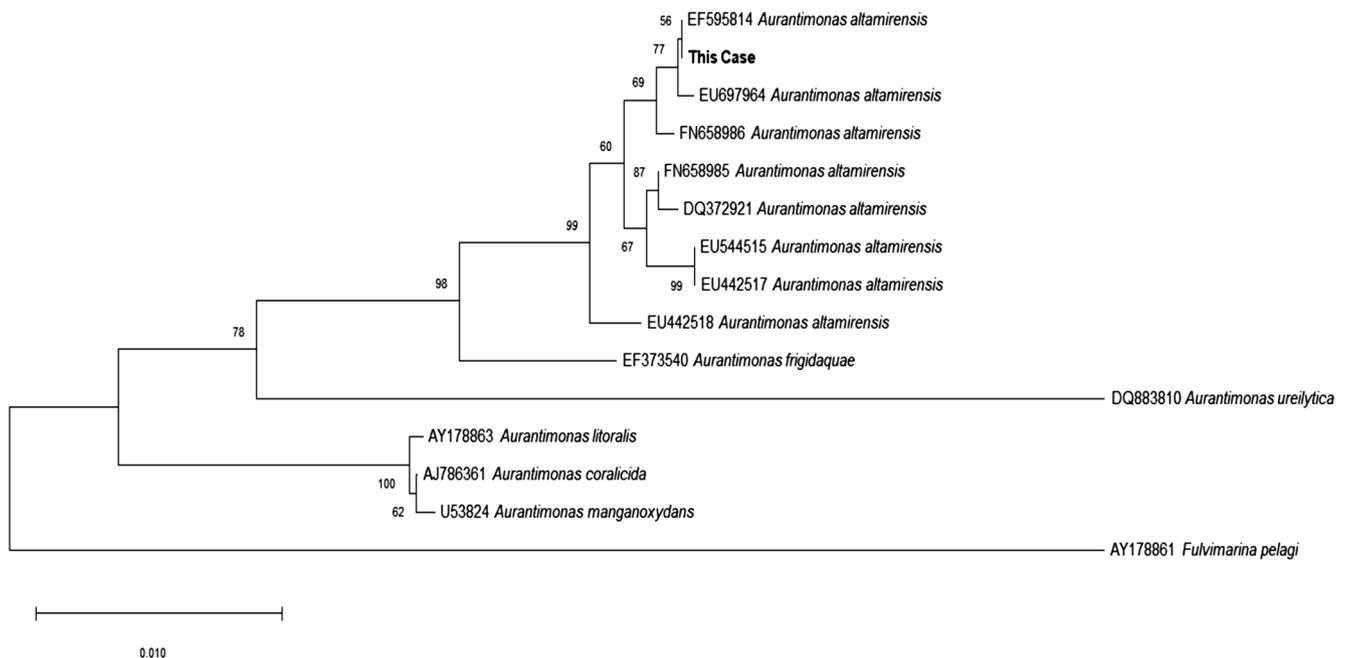
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**Table 1.** Invasive *Aureimonas altamirensis* infections in humans

Case No. (N)	Year reported	Age/Sex	Clinical diagnosis	Cultured sample	Comorbidity	Treatment	Outcome	Reference
1	2009	50/M	Soft tissue infection in scrotum	Blood	Diabetes mellitus, congestive heart failure, peripheral neuropathy, hyperthyroidism, hyperlipidemia, and chronic renal failure	Vancomycin, clindamycin, ciprofloxacin	Cured	4
2	2010	69/M	Parapneumonic effusion	Blood Pleural fluid	Chronic ischemic heart disease, total gastrectomy due to gastric cancer	Levofloxacin	Cured	5
3	2010	81/M	Parapneumonic effusion	Pleural fluid	NM	Levofloxacin	Cured	5
4	2012	65/F	Bacteremia	Blood	Multiple myeloma	Levofloxacin	Cured	6
5	2014	47/F	Peritonitis	Peritoneal fluid	Peritoneal carcinomatosis with liver and lymph node metastasis	Levofloxacin	Died	7
6	2015	NM	Cellulitis on forearm, wrist, and hand	Blood	NM	Vancomycin+imipenem, TMP-SMX+clindamycin	Cured	8

Abbreviations: M, male; F, female; NM, not mentioned; TMP-SMX, trimethoprim-sulfamethoxazole.



**Fig. 1.** Phylogenetic tree analysis based on 16S ribosomal RNA gene sequences. The tree was constructed using the neighbor-joining method with bootstrap resampling (1,000 replicates). The tree was rooted with *Fulvimarina pelagi* HTCC 2615 as the outgroup. Bootstrap values are expressed as percentages of 1,000 replications, and bars represent 0.010 substitutions per nucleotide position. GenBank database accession numbers are provided near the species names.

difference with the second most highly matched organism, *Aureimonas frigidaquae* (GenBank, NR\_044195.1), was 98.4% (1,357/1,379). A phylogenetic tree analysis with representative

*Aurantimonas* strains downloaded from BLAST (<http://www.ncbi.nlm.nih.gov/blast>) was performed using MEGA X (<https://www.megasoftware.net>) based on the 16S ribosomal RNA se-

quences [5, 9]. The results showed that our isolate formed a clade with the other *A. altamirensis* strains (Fig. 1).

Intravenous moxifloxacin was empirically administered, and the patient's condition improved. We performed positron emission tomography-computed tomography with [<sup>18</sup>F]-fluorodeoxyglucose to investigate the possible causes of fever. However, the results were normal, and repeated physical examinations and imaging studies showed that the patient did not have any other infection. In addition, the follow-up blood cultures were all negative, and the patient showed a good clinical response to empirical antibiotics. Therefore, we believed that the patient had primary *A. altamirensis* bacteremia according to the clinical information obtained from 16S ribosomal RNA sequencing. The fever subsided approximately two weeks after antibiotic treatment, and the patient was discharged.

We performed an antimicrobial susceptibility testing using the E-test (bioMérieux, Marcy L'Étoile, France) after receiving the 16S ribosomal RNA sequencing result. The minimum inhibitory concentrations for amoxicillin/clavulanic acid, gentamicin, imipenem, and trimethoprim/sulfamethoxazole were 1/0.5, 2, 0.125, and >32/608 µg/mL, respectively. *A. altamirensis* was susceptible to amoxicillin/clavulanic acid and imipenem, according to the non-species related breakpoints specified by the European Committee on Antimicrobial Susceptibility Testing guidelines [10].

A PubMed Medline search identified six cases of invasive human infection by *A. altamirensis* from blood, pleural fluid, and peritoneal fluid. All patients clinically improved with antibiotics, and only one patient died because of underlying comorbidities [7].

To date, the clinical relevance of *A. altamirensis* remains uncertain [5]. However, based on previous reports and our study, it is difficult to negate its potential pathogenic significance in humans. Information on the susceptibility of *A. altamirensis* to most antibiotics is limited, and clinicians rely on only a few case reports and empirical treatments. However, most clinical isolates were susceptible to the major antibiotics used for gram-negative bacteria, except quinolone, and showed successful treatment responses [2, 4, 5, 7].

In conclusion, although there is limited information on human infections, *A. altamirensis* should be considered an opportunistic pathogen, especially in immunocompromised patients. Additionally, laboratory physicians should be careful when there are discrepancies between identification results from automated systems and biochemical tests with growth conditions.

## Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

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