



# Comparison of Three Blood Collection Tubes for 35 Biochemical Analytes: The Becton Dickinson Barricor Tube, Serum Separating Tube, and Plasma Separating Tube

Sunghwan Shin , M.D., Jongwon Oh , M.D., and Hyung-Doo Park , M.D., Ph.D.

Department of Laboratory Medicine and Genetics, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea

The Barricor tube (Becton Dickinson [BD], Sunnyvale, CA, USA) was recently developed to mechanically separate plasma by increasing the centrifugation rate. We compared the Barricor tube with existing serum- and plasma-based tubes based on 35 biochemical analytes and preanalytical turnaround time (TAT). Blood samples were collected from 30 healthy volunteers in a Barricor tube, serum separating tube (SST, Vacutainer SST II Tube 8.5 mL, #368972; BD), or plasma separating tube (PST, Vacutainer PST Tube 8.0 mL, #367964; BD) in random order. Next, 27 chemistry analytes, six immunochemistry analytes, and two cardiac markers were compared using Passing-Bablok regression and the Bland-Altman method. Preanalytical TAT was measured for each tube.

The Barricor tube exhibited bias exceeding the desirable limit for nine and four analytes compared with the SST and PST, respectively. The Barricor tube lactate dehydrogenase value showed a bias of -10.29% and -9.86% compared with that of the SST and PST, respectively. The preanalytical TAT of Barricor tube was 8.8 minutes, which was the shortest among the three tubes. The clinical performance of the Barricor tube was equivalent to that of the SST and PST for most analytes, with an apparent advantage in preanalytical TAT. When using the Barricor tube, the reference range needs to be changed for some analytes that exceed the desirable bias limit.

**Key Words:** Barricor tube, Bias, Turnaround time, Analytes, Serum separating tube, Plasma separating tube

**Received:** February 3, 2020

**Revision received:** March 5, 2020

**Accepted:** July 29, 2020

**Corresponding author:**

Hyung-Doo Park, M.D., Ph.D.

Department of Laboratory Medicine and Genetics Samsung Medical Center, Sungkyunkwan University School of Medicine, 81 Irwon-ro, Gangnam-gu, Seoul 06351, Korea

Tel: +82-2-3410-0290

Fax: +82-2-3410-2719

E-mail: nayadoo@hanmail.net



© Korean Society for Laboratory Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

The BD Vacutainer Barricor plasma blood collection tube (Becton Dickinson [BD], Sunnyvale, CA, USA) is a recently introduced plasma separating tube (PST) that utilizes a mechanical separator which allows a higher centrifugation speed. We measured a total of 35 analytes using the Barricor tube (Vacutainer Barricor Tube 5.5 mL, #365057; BD), serum separating tube (SST; Vacutainer SST II Tube 8.5 mL, #368972; BD), and PST (Vacutainer PST Tube 8.0 mL, #367964; BD), for performance evaluation of Barricor tubes. Previous studies on Barricor tube

were either performed only with PST [1-4] or without considering immunochemistry analytes when compared with SST [5]. Other than comparison studies on Barricor tube, centrifugation conditions and sample quality have also been studied [6]; furthermore, hemolytic index and processing under vacuum have also been explored [7]. This study provides comparison data for the Barricor tube, SST, and PST for 27 chemistry analytes and two cardiac markers and for the Barricor tube and SST for six immunochemistry analytes, thus covering more analytes than

before with SST [4, 5]. The preanalytical turnaround time (TAT) of the Barricor tube, SST, and PST was compared, and the current status of the SST sample quality in our routine chemistry laboratory was also assessed. A total of 30 healthy volunteers were recruited prospectively. All volunteers were over 19 years of age and none were pregnant or breastfeeding; 22 mL of blood was collected by venipuncture from all volunteers, after obtaining informed consent. This study was conducted at Samsung Medical Center, Seoul, Korea, in March 2018. The study was reviewed and approved by the Samsung Medical Center Institutional Review Board (IRB no. 2018-01-101).

For TAT assessment, each time point was recorded manually at the start of blood collection, before centrifugation, before measuring the analytes with sample aliquots, and at the final reporting time of the analyzed results from selected equipment. The TAT preanalytical phase, calculated as the sum of the coagulation time, centrifugation time, time for visual inspection, aliquotting time, and time spent in sending samples to the equipment, was determined for each tube. Samples in SSTs were allowed to clot for 30 minutes immediately after blood collection and were then centrifuged at 25°C and 2,000×g for 10 minutes, while those in Barricor tubes and PSTs were centrifuged immediately after blood collection at 1,300×g for 10 minutes and 3,500×g for 5 minutes, respectively, at 25°C. Although the optimal centrifugation condition for the Barricor tube provided by the manufacturer is 4,000–5,000×g for 3 minutes, different conditions were used owing to the equipment currently installed in the clinical chemistry laboratory at the Samsung Medical Center.

A total of 27 routine chemistry analytes were measured with a Roche Cobas c702 chemistry analyzer (Roche Diagnostics, Basel, Switzerland), using aliquots from the Barricor tubes, SSTs, and PSTs (Table 1). Six immunochemistry analytes were measured with an ADVIA Centaur XP Immunoassay System (Siemens Healthineers, Erlangen, Germany), using aliquots from the Barricor tubes and SSTs (Table 2). Cardiac markers, cardiac troponin I (cTnI) and creatine kinase-myocardial band (CK-MB), were measured with an ADVIA Centaur XP Immunoassay System (Siemens), using aliquots from the Barricor tubes, SSTs, and PSTs. The performance of the Barricor tube was compared with that of the SST and PST using Passing-Bablok regression. The slope and intercept of the regression line were calculated with 95% confidence interval (CI). Percent biases at medical decision points were calculated for analytes with either a Passing-Bablok slope of one outside the 95% CI or an intercept of zero outside the 95% CI (Table 3). Statistically significant differences between the Barricor tube and SST were calculated using paired t-test,

and  $P < 0.05$  was considered statistically significant. Clinical significance was judged based on the desirable biological variation database or total allowable error according to the Clinical Laboratory Improvement Amendments criteria, whichever was lower [8, 9].

Nine analytes from the Barricor tube exceeded the desirable bias limit compared with those from the SST. Compared with that for the SST, the lactate dehydrogenase (LDH) value for the Barricor tube showed a percent bias of -10.29%, which exceeded the desirable bias limit. The biases for albumin (ALB), aspartate aminotransferase (AST), calcium (Ca), glucose, potassium, phosphorus, sodium, and total protein also exceeded the desirable bias limit, ranging from -6.75% to 4.78%.

When the Barricor tube was compared with the PST, all analytes, except AST, Ca, carbon dioxide (CO<sub>2</sub>), and LDH, were within the desirable bias limit, and values exceeding the desirable bias limit ranged from -1.16% to -9.86%. The Ca bias was higher than the desirable bias limit compared with both SST and PST, although the value for both was < 1.5%. Of the immunochemistry analytes, the folate and free thyroxine (FT4) values for the Barricor tube exceeded the desirable bias limit compared with those for the SST (Table 2). Regression for cTnI was not calculated, as most of the measured data were identical for the Barricor tubes, SST, and PST at values of 0.006 µg/L, which is the limit of detection of the test.

SST sample quality was also evaluated in our routine chemistry laboratory. Throughout the observational period, from January 11–16, 2019, 3,057 SSTs in total were visually inspected, after centrifugation, for quality review. Employees counted the number of samples containing artifacts such as gel globules or fibrin; of the SSTs, 229 (7.5%) required manual removal of either gel globules or fibrin after the centrifugation process. This remediation rate was consistent with the results of a global survey, which reported a remediation incidence of up to 6.0% for gel globule or fibrin [10]. Barricor tubes with less gel globules or fibrin could thus reduce manual labor and preanalytical TAT. Issues related to fibrin interference are known, and some clinical laboratories prefer plasma to serum [11]. The Barricor tubes presented the shortest preanalytical TAT, with an average preanalytical TAT of 8.8 minutes, owing to the short centrifugation step and higher centrifuge speed, whereas the SSTs exhibited the longest preanalytical TAT (average=43.6 minutes) owing to a coagulation step of 30 minutes. The average preanalytical TAT of the PSTs was 13.8 minutes.

Studies on Barricor tubes in terms of analyte stability, effect of centrifugation speed, performance comparison with SST or PST,

Table 1. Analytical data for Barricor tubes, SSTs, and PSTs and comparison results

Analyte	Barricor		SST		PST		Barricor vs SST			Barricor vs PST			Desirable bias (%) <sup>†</sup>
	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)	% Bias	P-B 95% CI slope	P-B 95% CI intercept	P (SST)	% Bias	P-B 95% CI slope	P-B 95% CI intercept	P (PST)	
ALB (g/L)	46.0 (2.5)	46.9 (2.3)	46.4 (2.3)	46.4 (2.3)	-1.90	1.00-1.33	-1.60-0.10	<0.05	-0.97	1.00-1.20	-0.96-0.00	<0.05	1.43
ALP (µkat/L)	0.95 (0.22)	0.97 (0.22)	0.96 (0.22)	0.96 (0.22)	-2.34	0.96-1.00	-2.00-1.25	<0.05	-0.33	0.94-1.00	-1.00-3.59	0.50	6.72
ALT (µkat/L)	0.35 (0.45)	0.36 (0.45)	0.35 (0.45)	0.35 (0.45)	-7.53	0.89-1.01	-1.44-0.67	<0.05*	-1.83	0.88-1.02	-1.23-1.46	0.20*	11.48
AMY (µkat/L)	1.06 (0.42)	1.06 (0.42)	1.06 (0.43)	1.06 (0.43)	-0.53	0.99-1.03	-1.71-0.35	0.137	-0.35	0.97-1.00	-0.20-1.40	0.073	7.40
AST (µkat/L)	0.33 (0.25)	0.35 (0.25)	0.35 (0.25)	0.35 (0.25)	-6.75	0.95-1.33	-7.07-0.50	0.06*	-8.82	0.99-1.20	-5.43-0.84	<0.05*	6.54
BUN (mmol/L)	3.85 (1.15)	3.86 (1.10)	3.87 (1.15)	3.87 (1.15)	-0.72	1.00-1.07	-0.74-0.00	0.54	-0.83	1.00-1.07	-0.76-0.00	0.18	5.60
Ca (mmol/L)	2.33 (0.05)	2.36 (0.05)	2.35 (0.05)	2.35 (0.05)	-1.46	0.82-1.25	-2.51-1.58	<0.05	-1.16	1.00-1.50	-4.93-0.00	<0.05	0.80
CO <sub>2</sub> (mmol/L)	26.31 (2.79)	26.62 (2.52)	27.12 (2.30)	27.12 (2.30)	-1.26	0.77-1.30	-8.27-6.74	0.55	-3.23	0.93-1.40	-10.99-1.92	0.074	1.60
CL (mmol/L)	101 (1.64)	101.17 (1.81)	101.01 (1.53)	101.01 (1.53)	-0.16	0.75-1.00	0.00-25.5	0.42	0	1.00-1.34	-34.71-0.00	0.68	0.50
CHOL (mmol/L)	4.38 (0.60)	4.48 (0.60)	4.42 (0.60)	4.42 (0.60)	-2.24	0.95-1.06	-14.87-5.00	<0.05	-0.92	0.97-1.05	-9.61-4.13	<0.05	4.10
CREAT (µmol/L)	64.55 (12.14)	64.55 (14.51)	66.32 (12.38)	66.32 (12.38)	0.23	0.86-1.10	-0.07-0.10	0.912	-2.26	0.87-1.08	-0.08-0.08	<0.05	4.00
DBIL (µmol/L)	4.62 (1.86)	4.62 (1.86)	4.45 (2.79)	4.45 (2.79)	1.33	1.00-1.00	0.00-0.00	1.00*	4.89	1.00-1.00	0.00-0.00	0.52*	14.20
TBIL (µmol/L)	9.58 (4.71)	9.92 (4.71)	9.92 (4.86)	9.92 (4.86)	-1.90	1.00-1.00	0.00-0.00	0.10	-2.82	1.00-1.00	0.00-0.00	0.16	9.00
GGT (µkat/L)	0.35 (0.34)	0.36 (0.34)	0.36 (0.34)	0.36 (0.34)	-0.89	1.00-1.02	-0.62-0.00	0.58*	-2.32	0.90-1.01	-1.00-1.19	0.53*	11.10
Glucose (mmol/L)	5.35 (0.71)	5.09 (0.66)	5.32 (0.66)	5.32 (0.66)	4.78	0.94-1.17	-10.17-9.63	<0.05	0.49	0.98-1.17	-14.67-2.45	0.31	2.30
HDL (mmol/L)	1.67 (0.44)	1.69 (0.44)	1.68 (0.44)	1.68 (0.44)	-0.97	0.98-1.00	-1.00-0.74	<0.05	-0.44	1.00-1.02	-1.39-0.00	0.165	5.60
Fe (µmol/L)	16.31 (5.59)	16.84 (5.59)	16.59 (5.59)	16.59 (5.59)	-3.46	0.97-1.01	-4.15-0.59	<0.05	-1.96	1.00-1.00	-2.00-1.00	<0.05	8.80
LDL (mmol/L)	2.79 (0.48.9)	2.83 (0.49)	2.78 (0.49)	2.78 (0.49)	-1.36	0.96-1.11	-12.67-3.15	<0.05	0.24	0.96-1.07	-7.04-5.00	0.596	5.46
LDH (µkat/L)	2.82 (0.71)	3.12 (0.73)	3.10 (0.72)	3.10 (0.72)	-10.29	0.90-1.35	-74.32-1.95	<0.05	-9.86	0.92-1.28	-62.38-1.42	<0.05	4.30
LIP (µkat/L)	0.56 (0.74)	0.57 (0.74)	0.56 (0.74)	0.56 (0.74)	-0.95	0.95-1.01	-0.45-0.97	0.136*	0.62	0.95-1.02	-0.49-1.17	0.309*	11.30
Mg (mmol/L)	0.83 (0.05)	0.84 (0.05)	0.84 (0.05)	0.84 (0.05)	-1.47	1.00-1.00	-0.05-0.00	<0.05	-1.50	1.00-1.00	0.00-0.00	<0.05	1.80
Potassium (mmol/L)	4.07 (0.23)	4.25 (0.24)	4.04 (0.24)	4.04 (0.24)	-4.41	0.67-1.25	-1.33-1.30	<0.05	0.75	0.80-1.50	-1.90-0.77	0.43	1.80
Phosphorus (mmol/L)	1.09 (0.14)	1.15 (0.14)	1.08 (0.14)	1.08 (0.14)	-5.69	1.00-1.00	-0.20-0.20	<0.05	0.31	1.00-1.00	0.00-0.00	0.33	3.38
Na (mmol/L)	140.07 (1.89)	140.37 (1.81)	139.93 (1.80)	139.93 (1.80)	-0.21	1.00-1.50	-71.25-0.00	0.184	0.00	1.00-1.00	0.00-0.00	0.46	0.20
TP (g/L)	74.00 (3.7)	72.60 (3.5)	74.90 (3.3)	74.90 (3.3)	1.94	0.83-1.33	-2.35-1.44	<0.05	-1.20	0.90-1.50	-3.85-0.64	<0.05	1.40
TG (mmol/L)	1.13 (0.48)	1.14 (0.50)	1.11 (0.49)	1.11 (0.49)	-1.12	0.93-1.00	-1.00-3.75	0.060	1.83	0.96-1.00	0.00-4.19	<0.05	9.60
UA (mmol/L)	0.29 (0.08)	0.29 (0.08)	0.29 (0.08)	0.29 (0.08)	0.27	1.00-1.06	-0.24-0.10	0.28	0.23	1.00-1.02	-0.08-0.00	0.33	4.90
cTnI (µg/L)	0.007 (0.006)	0.007 (0.006)	0.007 (0.005)	0.007 (0.005)	0.27	N/A <sup>†</sup>	N/A <sup>†</sup>	1.00	1.00*	N/A	N/A	1.00*	16.32
CK-MB (µg/L)	0.99 (0.82)	0.97 (0.79)	0.98 (0.81)	0.98 (0.81)	2.00	0.95-1.14	-0.09-0.07	0.20	1.78	0.93-1.06	-0.06-0.08	0.57	14.88

Measured with Roche Cobas c702 chemistry analyzer (Roche Diagnostics, Basel, Switzerland) except for cTnI and CK-MB which were measured with ADVIA Centaur XP Immunoassay System (Siemens Healthineers, Erlangen, Germany). Bold numbers indicate percent bias exceeding the desirable bias limit or Passing-Bablok intercept of zero outside the 95% CI.  
<sup>\*</sup> Calculated with paired Wilcoxon signed-rank test. All other values without asterisks were calculated with paired t-test; †Regression was not calculated; ‡Desirable bias provided in the desirable biological variation database specifications [8].  
Abbreviations: ALB, albumin; ALT, alanine aminotransferase; ALP, alkaline phosphatase; AMY, amylase; AST, aspartate aminotransferase; BUN, blood urea nitrogen; Ca, calcium; CO<sub>2</sub>, carbon dioxide; CL, chloride; CHOL, cholesterol; CREAT, creatinine; DBIL, direct bilirubin; TBIL, total bilirubin; GGT, γ-glutamyltransferase; HDL, high density lipoprotein; Fe, iron; LDL, low density lipoprotein; LDH, lactate dehydrogenase; LIP, lipase; Mg, magnesium; Na, sodium; TP, total protein; TG, triglycerides; UA, uric acid; cTnI, cardiac troponin I; CK-MB, creatine kinase-MB; SST, serum separating tube; PST, plasma separating tube.

**Table 2.** Analytical data for Barricor tube and SST immunochemistry analyte values

Analyte	Barricor	SST	%Bias	Barricor vs SST			Desirable bias (%) <sup>‡</sup>
	Mean (SD)	Mean (SD)		Passing-Bablok 95% CI slope	Passing-Bablok 95% CI intercept	P (SST)	
Vit. B <sub>12</sub> (pmol/L)	444.72 (186.90)	400.42 (161.28)	10.01	<b>1.06–1.20</b>	-45.99–18.83	<0.05	17.70
Folate (nmol/L*)	31.77 (8.35)	20.58 (6.88)	45.13	0.98–1.17	3.26–4.86	<0.05	19.20
Ferritin (µg/L)	96.10 (108.45)	106.3 (115.5)	-1.48	<b>0.91–0.97</b>	-0.57–1.69	<0.05 <sup>†</sup>	5.20
TSH (mIU/L*)	1.54 (0.97)	1.54 (0.96)	-3.54	0.98–1.02	-0.02–0.03	0.88	7.80
FT4 (pmol/L*)	16.86 (3.33)	15.96 (3.68)	5.31	0.84–1.29	-0.27–0.26	<0.05	3.30
FSH (IU/L*)	12.38 (21.07)	12.40 (22.09)	3.16	0.94–1.07	-0.21–0.47	0.012 <sup>†</sup>	12.10

Measured with ADVIA Centaur XP Immunoassay System (Siemens Healthineers, Erlangen, Germany). Bold numbers indicate Passing-Bablok slope of one outside the 95% CI.

\*Reagent was not validated for plasma; thus, the Barricor tube results are unreliable; <sup>†</sup>Calculated with paired Wilcoxon signed-rank test. All other values were calculated with paired t-test; <sup>‡</sup>Desirable bias provided in the desirable biological variation database specifications [8].

Abbreviations: CI, confidence interval; Vit. B<sub>12</sub>, vitamin B<sub>12</sub>; TSH, thyroid stimulating hormone; FT4, free thyroxine; FSH, follicle-stimulating hormone; SST, serum separating tube.

**Table 3.** Percentage bias at medical decision points for selected analytes in Barricor tube

Analyte	Barricor tube						Desirable Bias (%) <sup>†</sup>
	Medical decision point 1	% Bias (vs SST)	% Bias (vs PST)	Medical decision point 2	% Bias (vs SST)	% Bias (vs PST)	
ALB (g/L)	35	<b>-2.86</b>	-1.27	52	<b>-1.92</b>	<b>-1.71</b>	1.43
AST (µkat/L)	0.53	-2.52	-6.25	1.67	0.04	-2.00	6.54
Fe (µmol/L)	5.91	<b>-9.10</b>	-6.06	34.55	-1.55	-1.04	8.80
LDH (µkat/L)	2.25	<b>-12.64</b>	<b>-6.55</b>	3.57	<b>-14.24</b>	<b>-6.40</b>	5.46
Phosphorus (mmol/L)	0.81	<b>-8.00</b>	0.00	1.45	<b>-4.44</b>	0.0	3.38
Vit B <sub>12</sub> (pmol/L)	155.72	6.44	N/A*	673.32	10.68	N/A*	17.70
Ferritin (µg/L)	10	-4.79	N/A*	291	<b>-6.82</b>	N/A*	5.20

Bold numbers indicate percent bias exceeding the desirable bias limit.

\*Comparison between Barricor tube and PST was not performed; <sup>†</sup>Desirable bias provided in the desirable biological variation database specifications [8].

Abbreviations: SST, serum separating tube; PST, plasma separating tube; ALB, albumin; AST, aspartate aminotransferase; LDH, lactate dehydrogenase; Fe, iron; Vit. B<sub>12</sub>, vitamin B<sub>12</sub>.

and effects on therapeutic drug monitoring have reported stable performance of Barricor tubes for most routine chemistry tests [1–7, 12]. While most analytes included in Barricor tube comparison studies have yielded clinically non-significant results, LDH has been commonly reported to have a bias compared with SST or PST [1, 3–5]. Our Barricor tube LDH values showed a bias of ~10% compared with those obtained using SST or PST. Although Arslan, *et al.* [5] reported a positive bias of 10–20% using Barricor tubes compared with SSTs and PSTs, our LDH results showed a negative bias. Previous studies have reported LDH values with a positive or negative bias <5% compared with those using PSTs [1, 3, 4]. In our study, AST using Barricor tubes showed a negative bias >5% compared with that using SST, and the value exceeded the desirable bias limit. Potassium and phosphorus using Barricor tubes showed a negative bias close to 5%, and the value

exceeded the desirable bias limit compared with those using SST.

AST, alanine aminotransferase, and LDH, along with many other analytes, are known to be affected by hemolytic interference [13], which is suspected as the cause of discrepant results. The higher centrifuge speed required for the Barricor tube might cause hemolytic interference. However, studies on the effects of centrifuge speed on the Barricor tube for measured analytes have not provided meaningful results [4, 5]. Several stability studies have suggested prolonged contact of serum/plasma and clot along with disrupted cell-membrane integrity of erythrocytes as factors contributing to the increased LDH values using serum or plasma [11, 14, 15]. LDH value is also known to increase with membrane leakage from necrotic cells [16]. Another stability study with Barricor tubes showed more stable LDH values compared with PSTs [1]. The Barricor tube mechanical sep-

arators provide cell and plasma separation using a different mechanism from that of gel separators, and this difference could contribute to varying sample separation quality, resulting in different LDH values. In cases of hemolytic interference, the biases observed for AST and LDH should be either both positive or both negative, as both are intracellular components released during hemolysis [13]. While our study using Barricor tubes showed negative biases for both AST and LDH, Arslan, *et al.* [5] reported positive biases for both AST and LDH. The different experimental conditions used in the two studies might have led to a different hemolytic effect. However, further studies with varying centrifugation speed, temperature, and aliquot conditions are required to elucidate these discrepancies.

Potassium values are known to be elevated in serum, and our results are consistent with previous findings [17]. In addition, the potassium values using the Barricor tubes were comparable to those using PSTs, and this result is consistent with previous findings [1, 3–5]. Glucose value has been reported to be lower in serum, as the 30-minute coagulation step required for SST increases the chance of glycolysis during the process [18, 19].

Of the six immunochemistry analytes tested, folate and FT4 showed biases exceeding the desirable bias limit (45.13% and 5.31%, respectively). While the reagents used for the immunochemistry analytes vitamin B<sub>12</sub> (Vit B<sub>12</sub>) and ferritin were validated for both serum and plasma, the reagents for folate, thyroid stimulating hormone (TSH), FT4, and follicle-stimulating hormone (FSH) were only validated for serum samples. The measured bias of Vit. B<sub>12</sub> was 10.01%, which was lower than the desirable bias limit. A previous study reported that Vit. B<sub>12</sub> values using the Barricor tubes were clinically equivalent to those using SSTs and PSTs, with a bias <1% [5]. Fournier, *et al.* [1] reported a bias of 0.08% for Vit. B<sub>12</sub> in a comparative study between the Barricor tubes and PSTs; however, they did not perform a comparison with SSTs.

To date, the Barricor tubes and SSTs have not been compared with respect to the immunochemistry analytes folate, ferritin, TSH, FT4, and FSH. Of these six immunochemistry analytes, our data showed that Vit. B<sub>12</sub>, ferritin, TSH, and FSH were within the desirable bias limit. Although the reagents for TSH and FSH were not validated for plasma, the Barricor tube-measured data were still within the desirable bias limit.

Of the selected analytes, the LDH bias exceeded the desirable bias limit at the medical decision points, compared with those of both SST and PST, with biases ranging from -6.40% to -14.24% (Table 3). In addition, the biases for ALB, Fe, phosphorus, and ferritin were higher than the desirable bias limit at one or more

of the medical decision points.

As this study used peripheral blood samples from healthy volunteers, we could not conduct a performance evaluation covering the entire analytical measurement range. In addition, most of the cTnI values showed limit values <0.006 µg/L. These values were not normally distributed, which was a limiting factor of this study.

In conclusion, the clinical performance of Barricor tubes was equivalent to that of SSTs and PSTs for most of the analytes. AST, Ca, and LDH showed relatively high biases, exceeding the desirable bias limit. Vit. B<sub>12</sub> also showed a relatively high bias using the Barricor tube compared with SST; however, the value was still within the acceptable range. We showed clinically equivalent performance for the Barricor tubes at a centrifugation speed of 3,500×g, which was slower than the optimal speed recommended by the manufacturer. When using the Barricor tube, the reference range might need to be changed for analytes exceeding the desirable bias limit. Moreover, additional studies on the Barricor tube are needed to elucidate the discrepant biases between our study and previous studies.

## ACKNOWLEDGEMENTS

We thank BD for providing Barricor tubes for this study.

## AUTHOR CONTRIBUTIONS

HDP designed the study concept and reviewed data analysis and manuscript. JO reviewed the data. SS performed statistical data analysis and wrote the manuscript. All the authors have accepted responsibility for the entire content of this submitted manuscript and approved submission.

## CONFLICTS OF INTEREST

None declared.

## RESEARCH FUNDING

None declared.

## ORCID

Sunghwan Shin <https://orcid.org/0000-0003-1038-029X>  
Jongwon Oh <https://orcid.org/0000-0002-6967-7365>  
Hyung-Doo Park <https://orcid.org/0000-0003-1798-773X>

## REFERENCES

1. Fournier JE, Northrup V, Clark C, Fraser J, Howlett M, Atkinson P, et al. Evaluation of BD Vacutainer Barricor blood collection tubes for routine chemistry testing on a Roche Cobas 8000 Platform. *Clin Biochem* 2018; 58:94-9.
2. Füzéry AK, Raizman JE, Goudreau BL, Moses K, Niemann K, Park J, et al. The BD Barricor blood collection tube is an acceptable and robust alternative to the PST for use with the Beckman AccuTnl+3 assay. *Clin Biochem* 2017;50:851-7.
3. Dupuy AM, Badiou S, Daubin D, Bargnoux AS, Magnan C, Klouche K, et al. Comparison of Barricor vs. lithium heparin tubes for selected routine biochemical analytes and evaluation of post centrifugation stability. *Biochem Med (Zagreb)* 2018;28:020902.
4. Cadamuro J, Mrazek C, Leichtle AB, Kipman U, Felder TK, Wiedemann H, et al. Influence of centrifugation conditions on the results of 77 routine clinical chemistry analytes using standard vacuum blood collection tubes and the new BD-Barricor tubes. *Biochem Med (Zagreb)* 2018;28:010704.
5. Arslan FD, Karakoyun I, Basok BI, Aksit MZ, Baysoy A, Ozturk YK, et al. The local clinical validation of a new lithium heparin tube with a barrier: BD Vacutainer Barricor LH Plasma tube. *Biochem Med (Zagreb)* 2017; 27:030706.
6. Padoan A, Zaninotto M, Piva E, Sciacovelli L, Aita A, Tasinato A, et al. Quality of plasma samples and BD Vacutainer Barricor tubes: effects of centrifugation. *Clin Chim Acta* 2018;483:271-4.
7. Ramakers C. BD Vacutainer Barricor tube in the emergency department: reduced hemolysis rates using partial draw tubes with reduced vacuum. *Clin Chem Lab Med* 2018;56:e31-2.
8. Desirable specifications for total Error, imprecision, and bias, derived from intra- and inter-individual biologic variation. <https://www.westgard.com/biodatabase1.htm>. accessed August 1st 2019.
9. Clinical laboratory improvement amendments of 1988: final rule. Department of Health and Human Services, Health Care Financing Administration; Federal Register 57, no. 7146 (1992).
10. Erdal EP, Mitra D, Khangulov VS, Church S, Plokhoy E. The economic impact of poor sample quality in clinical chemistry laboratories: results from a global survey. *Ann Clin Biochem* 2017;54:230-9.
11. Boyanton BL Jr. and Blick KE. Stability studies of twenty-four analytes in human plasma and serum. *Clin Chem* 2002;48:2242-7.
12. Schrapp A, Mory C, Duflo T, Pereira T, Imbert L, Lamoureux F. The right blood collection tube for therapeutic drug monitoring and toxicology screening procedures: standard tubes, gel or mechanical separator? *Clin Chim Acta* 2019;488:196-201.
13. Lippi G, Salvagno GL, Montagnana M, Brocco G, Guidi GC. Influence of hemolysis on routine clinical chemistry testing. *Clin Chem Lab Med* 2006; 44:311-6.
14. Zhang DJ, Elswick RK, Miller WG, Bailey JL. Effect of serum-clot contact time on clinical chemistry laboratory results. *Clin Chem* 1998;44: 1325-33.
15. Monneret D, Godmer A, Le Guen R, Bravetti C, Emerald C, Marteau A, et al. Stability of routine biochemical analytes in whole blood and plasma from lithium heparin gel tubes during 6-hr storage. *J Clin Lab Anal* 2016;30:602-9.
16. Chan FK, Moriwaki K, De Rosa MJ. Detection of necrosis by release of lactate dehydrogenase activity. *Methods Mol Biol* 2013;979:65-70.
17. Bowen RA, Hortin GL, Csako G, Otañez OH, Remaley AT. Impact of blood collection devices on clinical chemistry assays. *Clin Biochem* 2010;43: 4-25.
18. Dimeski G, Yow KS, Brown NN. What is the most suitable blood collection tube for glucose estimation? *Ann Clin Biochem* 2015;52:270-5.
19. Lippi G, Salvagno GL, Lampus S, Danese E, Gelati M, Bovo C, et al. Impact of blood cell counts and volumes on glucose concentration in uncentrifuged serum and lithium-heparin blood tubes. *Clin Chem Lab Med* 2018;56:2125-31.